Effect of Withinia Somnifera and Shilajit on Alcohol Addiction in Mice

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ABSTRACT
Background: Alcohol addiction is a social problem leading to both loss of health and economic prosperity among addicted individuals. Common properties of anti-addictive compounds include anti-anxiety, anxiolytics, anxiety, sedative, and nootropics actions primarily through modulation of gamma-aminobutyric acid (GABA) and serotonergic systems. Objective: Here, we screen ashwagandha and shilajit known ethnopharmacologically as nervine tonic and adaptogenic herbs for possible anti-addictive potential. Materials and Methods: Effect of ashwagandha churna and shilajit was measured on ethanol withdrawal anxiety using elevated plus maze. Effect of ashwagandha and shilajit was also measured using two bottle choice protocol of voluntary drinking. Results: Both ashwagandha and shilajit were found to reduce alcohol withdrawal anxiety in a dose-dependent manner. These herbs alone or in combination also decreased ethanol intake and increased water intake significantly after 21 days of chronic administration. Chronic administration of ashwagandha was found to significantly increase GABA and serotonin levels whereas shilajit altered corticohippocampal dopamine in mice. Conclusion: These central nervous system active herbs alone or in combination reduced both alcohol dependence and withdrawal thus showing promising anti-addictive potential.

Key words: Addiction, alcohol, ashwagandha, mice, shilajit, withdrawal

SUMMARY
• Withania Somnifera alone and in combination with Shilajeet prevented ethanol withdrawal and alcohol addiction

INTRODUCTION
Alcohol addiction is a worldwide problem. In 2012, 3.3 million deaths have been reported due to alcohol consumption and is the fifth leading risk factor for premature death and disability. Alcohol contributes to over 200 diseases and injury-related health conditions, most notably alcohol dependence, liver cirrhosis, cancers, and injuries.[1,2] Alcohol dependence is a complex and dynamic process. Many neurobiological and environmental factors influence drinking habits.[3] An individual's tendency to imbibe to alcohol is primarily a balance between alcohol's rewarding effects and its negative consequences like withdrawal symptoms. Association of these good/bad feelings with environmental factors influence drinking habits.[3] Among its various physiological actions, its action on central nervous system (CNS) primarily involves its effect against cognitive impairment, anxiety, and depression.[4,5] Due to its anti-oxidant,[6] anti-inflammatory,[7] and neuroprotective effects,[8] Ashwagandha (Withania somnifera, family. Solanaceae) is commonly known as “Indian Winter cherry” or “Indian Ginseng.” Among its various physiological actions, its action on central nervous system (CNS) primarily involves its effect against cognitive impairment, anxiety, and depression.[4,5] Due to its anti-oxidant,[6] anti-inflammatory,[7] and neuroprotective effects,[8] Ashwagandha is predominantly used to improve cognitive function and reduce anxiety and depression. Shilajit is a blackish-brown powder or an exudate from high mountain rocks, which is commonly used in traditional medicine to improve cognitive function and reduce anxiety and depression.
especially found in the Himalayan Mountains between India and Nepal. Shilajit has been known and used for centuries in Ayurvedic medicine, as a rejuvenator and as an anti-aging compound. [23,24] Considering its unique composition as a phytocomplex, very rich in fulvic acid, researchers hypothesize that Shilajit is produced by the decomposition of plant material from species such as Euphorbia royleana and Trifolium repens. [19,25] Various pharmacological properties for shilajit including anti-inflammatory, [26] anti-oxidant, [27] immunomodulatory, [28] aphrodisiac [29] are well documented. In the CNS, it primarily acts as a memory enhancer, [30] neuroprotective, [31] and anxiolytic [32] agent. Shilajit has also been shown to have parasympathetic and GABA-mimetic actions. [33,34] Ashwagandha and a polyherbal formulation NR-ANX-C containing ashwagandha and shilajit may prevent alcohol-induced withdrawal anxiety in rats. [35,36] Here, we report that acute administration of ashwagandha, shilajit, and their combination may prevent alcohol withdrawal anxiety whereas their chronic administration (15 days) may reduce alcohol consumption in mice. Chronic administration of these herbal extracts may also alter cortical GABA, 5-HT, and dopamine levels, which may be responsible for their anti-addictive properties.

MATERIALS AND METHODS

Animals

Swiss albino mice (20–30 g) were used in this study. Animals were issued from the Institutional Animal House (Reg. No. 621/02/AC/CPCSEA) of Birla Institute of Technology, Mesra. All animals were kept in polycrylic cages and maintained under standard conditions (room temperature 24–27°C and humidity 60–65% with 12:12 lightdark cycles). The food was provided in the form of dry pellets and water ad libitum. The animals were allowed to get acclimatized to the laboratory conditions for 7 days before the commencement of the experiment. All experiments involving animals complied with the ethical standards of animal handling and approved by the Institutional Animal Ethics Committee (BIT/PH/IAEC/28/2013).

Estimation of blood alcohol levels

Blood was collected by retro-orbital bleeding with animals in light ether anesthesia after 20 min of ethanol administration. Ethanol levels were measured using ultraviolet (UV) assay kit for alcohol estimation based on manufacturer’s protocol (NZY Tech Genes and Enzymes Portugal).

Development of conditioned place preference model

Apparatus

The conditioned place preference (CPP) apparatus contain three compartments. The two end compartments (30.5 cm × 26.5 cm × 37 cm) were connected by a central corridor (12.75 cm × 23 cm × 15.25 cm). The compartment on the left had black walls with a perforated stainless steel floor with round holes on staggered centers. The central corridor was transparent with a smooth plexiglass floor, and the right compartment had white walls with a stainless steel mesh floor.

CPP was performed as described [25,26] with slight modifications. It mainly consists of three phases:

1. Preconditioning phase: (1st–2nd day) The animals were placed in the middle chamber and allowed to explore both the chambers for 30 min
2. Conditioning phase: (3rd–10th day) Each mouse was treated for eight consecutive sessions with alternate oral administration of alcohol and saline. On day 3, 5, 7, and 9, the animals were administered ethanol (2 g/kg b.w.; i.p. 10% [v/v]) and placed in one compartment for 30 min. In addition, on day 4, 6, 8, and 10, the animals were administered saline and placed in opposite compartment
3. Postconditioning phase: (11th–12th day) Mice were placed in the middle chamber and allowed free access to both chamber for 30 min.

Time spent in ethanol and saline-paired chamber was measured

4. Treatment protocol: After development of withdrawal (15th day), the following treatment schedule was followed:

- **Group 1**: Saline
- **Group 2**: Ethanol (2 g/kg, i.p. 10% [v/v] Bengal Chemical, Kolkata)
- **Group 3**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by ashwagandha churna (50 mg/kg, b.w., oral in 2% Tween 80; Withania somnifera root extracts; Vyas Pharmaceuticals, Haridwar) on the 15th day
- **Group 4**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by ashwagandha churna (100 mg/kg, oral in 2% Tween 80) on the 15th day
- **Group 5**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by ashwagandha churna (200 mg/kg, oral in 2% Tween 80) on the 15th day
- **Group 6**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by ashwagandha churna (500 mg/kg, oral in 2% Tween 80) on the 15th day
- **Group 7**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by shilajit (10 mg/kg, oral in 2% Tween 80; Shuddha Shilajit, Baidyanath, Kolkata) on the 15th day
- **Group 8**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by shilajit (25 mg/kg, oral in 2% Tween 80) on the 15th day
- **Group 9**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by shilajit (50 mg/kg, oral in 2% Tween 80) on the 15th day
- **Group 10**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by ashwagandha churna (500 mg/kg, oral) + shilajit (25 mg/kg, oral) on the 15th day
- **Group 11**: Ethanol (2 g/kg, i.p. 10% [v/v]) till day 10 followed by diazepam (1 mg/kg, i.p.) on the 15th day.

60 min after oral drug administration and 30 min after intraperitoneal administration, the behavioral tests were performed.

Behavioral studies to measure alcohol withdrawal anxiety

Elevated plus maze

Model has been validated pharmacologically and currently considered the “gold standard” test of anxiety-related behavior. Elevated plus maze (EPM) was performed as described by Kokare et al. [27] In summary, after drug treatment, individual mice was placed at the center of the maze, head facing an open arm. During the 5 min test period, the number of entries and time spent on the open arm were recorded automatically (Medicraft Electromedicals, Lucknow, India).

Chronic-treatment study to measure alcohol intake

Two-bottle choice ethanol drinking

We used the standard two-bottle choice protocol, which is a widely used animal model to capture aspects of voluntary alcohol consumption in humans. [38] Following 7 days of acclimatization, animals were subjected to an ethanol drinking acquisition regimen. The animals remained in their home cages at all times throughout the study but had their water bottles removed during a 4 h and ethanol presentation period. During this period, animals were exposed to a free choice between ethanol (15% v/v) and water for 20 days but with no drug pretreatment.

After 20 days of ethanol administration, animals were divided into different groups for 10 days of treatment. Each day, the bottles were weighed before and after 4 h of limited access period and the differences were used to calculate the water and ethanol intake. The mean intake was expressed as g/kg body weight/day of water and g/kg body weight/day of ethanol intake. All animals were given unrestricted
After 20 days of pretreatment with ethanol (15% v/v), the animals were divided into different treatment groups (n = 7/group) as follows:

- **Group 1:** (Control) received only water
- **Group 2:** (Positive control) received free choice ethanol (15%v/v)/water
- **Group 3:** Received free choice ethanol (15%v/v)/water and ashwagandha (500 mg/kg oral) 21st–30th day
- **Group 4:** Received free choice ethanol (15%v/v)/water and shilajit (25 mg/kg oral) 21st–30th day
- **Group 5:** Received free choice ethanol (15%v/v)/water and combination (ashwagandha 500 mg/kg, oral + shilajit 25 mg/kg oral) 21st–30th day
- **Group 6:** Received free choice ethanol (15%v/v)/water and diazepam (10 mg/kg oral) 21st–30th day.

After the above experimental protocol of 30 days, 4 animals/group were sacrificed under ether anesthesia by cervical dislocation for biochemical estimation.

**Estimation of gamma-aminobutyric acid levels from brain tissue**

Brain tissue was homogenized in 5 ml of 0.01 M HCl. In this homogenate, 8 ml of ice cold ethanol was added and kept for 1 h at 0°C. The contents were centrifuged for 10 min at 16,000 rpm and supernatant was collected in a Petridish. The precipitate was washed 3 times with 5 ml of 75% ethanol. The washes were combined with supernatant and evaporated to dryness. To the dry mass 1 ml water and 2 ml chloroform were added and centrifuged at 2000 rpm. Upper phase containing GABA was separated, and 10 μl of it was applied as spot on Whatman filter paper. The mobile phase consisted of n-butanol, acetic acid, water in 4:1:5 ratios. The chamber was saturated for half an hour with mobile phase. The paper chromatogram was developed with ascending technique. The paper was dried in a hot air oven and then sprayed with 0.5% ninhydrin solution. The paper was dried. Blue color spot developed on paper which was cut and heated with 2 ml ninhydrin solution on water bath at 60–65°C. Water was added to the solution and kept for 1 h and supernatant was used. Absorbance was measured at 570 nm on UV-visible spectrophotometer.

**Determination of DA and 5-HT**

Corticolimic slices were homogenized in 600 μL of ice cold solution of 0.4 M perchorlic acid containing 0.4 mM sodium metabisulfite and disodium ethylenediaminetetraacetic acid (EDTA), and centrifuged at 5000 g for 20 min at 4°C; supernatants were filtered through 0.45 μm cellulose membranes. The monoamine content (DA, 5-HT) of the brain tissue was determined using HPLC systems. The mobile phase consisted of n-butanol, acetic acid, water in 4:1:5 ratios. The chamber was saturated for half an hour with mobile phase. The paper chromatogram was developed with ascending technique. The paper was dried, and 10 μl of it was applied as spot on Whatman filter paper. The mobile phase consisted of n-butanol, acetic acid, water in 4:1:5 ratios. The chamber was saturated for half an hour with mobile phase. The paper chromatogram was developed with ascending technique. The paper was dried.

**RESULTS**

**Conditioned place preference**

In CPP test, the animal’s choice to spend more time in either environment provides a direct measure of the conditioned reinforcing effect of a drug. In our study, on day 11, control group (saline-treated animals) showed little increase in the preference for the ethanol-paired chamber as compared to the saline-paired chamber. However, ethanol-treated animals showed a significant increase in preference for the ethanol-paired chamber as compared to the saline-paired chamber. Ashwagandha also increased the number of open arm entries over 5 days ethanol abstinent animals, thus confirming its action against ethanol-abstinent anxiety [Figure 2].

**Effect of ashwagandha on withdrawal anxiety**

Ethanol consumption lead to increase in time spent in open arm of EPM in mice due to anxiolytic action of ethanol (data not shown). Five days postabstinence animals showed a significant decrease in time spent in the open arm of EPM as compared to animals on ethanol suggesting withdrawal anxiety. Ashwagandha led to significant increase in time spent in the open arm in a dose-dependent manner (50 mg/kg P < 0.01, 100 mg/kg P < 0.001, 200 mg/kg P < 0.001, and 500 mg/kg P < 0.001, n = 7/dose) thus suggesting reversal of ethanol withdrawal anxiety. Moreover, 200 mg/kg and 500 mg/kg treatment with ashwagandha showed a significant increase in time spent in open arm (P < 0.01 and P < 0.001, respectively; n = 7) compared to animals on ethanol suggesting intrinsic and more potent anxiolytic action of ashwagandha over ethanol. At higher doses of 200 and 500 mg/kg, ashwagandha also increased (P < 0.001; n = 7) the number of open arm entries over 5 days ethanol abstinent animals, thus confirming its action against ethanol-abstinent anxiety [Figure 2].

**Effect of shilajit on withdrawal anxiety**

Administration of shilajit to postabstinence animals led to significant increase in time spent in the open arm in a dose-dependent manner (P < 0.001 for 10 mg/kg, 25 mg/kg and 50 mg/kg; n = 7) thus reversing ethanol withdrawal anxiety. The time spent in the open arm was also significantly higher (P < 0.001) than animals on ethanol suggesting the intrinsic anxiolytic activity of shilajit. At doses of 25 and 50 mg/kg, shilajit also significantly increased (P < 0.01; n = 7) the number of entries in the open arm of EPM in ethanol abstinent animals [Figure 2]. Based on the above results, we studied the effect of optimal dose of ashwagandha (200 mg/kg), shilajit (25 mg/kg) and their combination on withdrawal anxiety. However, the combination of ashwagandha (200 mg/kg) and shilajit (25 mg/kg) did not show any significant increase in time spent...
Figure 2: Effect of ashwagandha and shilajit on ethanol withdrawal anxiety using elevated plus maze. (a) Ethanol abstinence significantly decreased \( (P < 0.001) \) time spend in open arm compared to ethanol-treated animals. Ashwagandha treatment to abstinent animals significantly (50 mg/kg \( P < 0.01 \) and 100 mg/kg, 200 mg/kg, 500 mg/kg \( P < 0.001 \)) increased the time spend in open arm compared to ethanol abstinent animals. (b) Ethanol abstinence significantly decreased \( (P < 0.001) \) the number of entries in open arm compared to animals on ethanol. Ashwagandha treatment (200 mg/kg and 500 mg/kg) significantly \( (P < 0.001) \) increased the number of open arm entries compared to abstinent animals. (c) Ethanol abstinence significantly decreased \( (P < 0.001) \) time spend in open arm compared to ethanol-treated animals. Shilajit treatment to abstinent animals significantly (10 mg/kg, 25 mg/kg, 50 mg/kg \( P < 0.001 \)) increased the time spend in open arm compared to ethanol abstinent animals. (d) Ethanol abstinence significantly decreased \( (P < 0.05) \) the number of entries in open arm compared to animals on ethanol. Shilajit treatment (25 mg/kg and 50 mg/kg) significantly \( (P < 0.01) \) increased the number of open arm entries compared to abstinent animals. Values (e) Ashwagandha (200 mg/kg) and shilajit (25 mg/kg) together significantly increased \( (P < 0.01) \) the time spend in open arm compared to ethanol abstinent animals. However, this increase was comparable \( (P > 0.05) \) with ashwagandha (200 mg/kg) and shilajit (25 mg/kg) treatments alone. Diazepam also significantly increased time spend in open arm over ethanol abstinent and ethanol-treated groups \( (P < 0.001) \). (f) Ashwagandha (200 mg/kg) and shilajit (25 mg/kg) together significantly increased \( (P < 0.01) \) the number of entries into the open arm when compared with ethanol abstinent animals. However, this increase was comparable \( (P > 0.05) \) with ashwagandha (200 mg/kg) and shilajit (25 mg/kg) treatments alone. Diazepam also significantly increased the number of open arm entries over ethanol abstinent and ethanol-treated groups \( (P < 0.001) \). Values represent mean ± standard error of the mean \( n = 7 \)
or the number of entries in the open arm ($P > 0.05$, $n = 7$) as compared to ashwagandha or shilajit alone in ethanol-withdrawn animals [Figure 2].

**Effect of ashwagandha and Shilajit on alcohol consumption**

Both ashwagandha (500 mg/kg) and shilajit (50 mg/kg) and their combination treated animals showed a significant ($P < 0.001$, $n = 7$) decrease in ethanol intake and a significant increase in ($P < 0.001$, $n = 7$) water intake as compared to control group after the 28th day or after 8 days of therapy. This was comparable with diazepam-treated animals which also showed a significant decrease ($P < 0.001$, $n = 7$) in ethanol intake and an increase in water intake after day 28 compared to control group [Figure 3]. Hence, we conclude that both ashwagandha and shilajit significantly decreased ethanol intake in mice.

**Effect of ashwagandha and shilajit on corticohippocampal monoamine levels**

Animals on free access to alcohol for 30 days showed a significant increase in serotonin as well as dopamine ($P < 0.05$, $n = 5$) compared to control animals. While animals with free access to ethanol (30 days) and receiving ashwagandha (500 mg/kg) or ashwagandha (500 mg/kg) and shilajit (50 mg/kg) for the last 10 days of this 30 days period showed a significant ($P < 0.01$, $n = 5$) increase in corticohippocampal serotonin compared to animals on ethanol (30 days). While shilajit treated animals did not show any significant increase in serotonin levels. However, treatment with shilajit (50 mg/kg) as well as ashwagandha and shilajit combination leads to a significant increase ($P < 0.01$) in dopamine levels compared to animals on ethanol (30 days). Diazepam treatment had little effect on dopamine and serotonin levels [Figure 4].

**Effect of ashwagandha and shilajit on gamma-aminobutyric acid levels**

After 30 days of alcohol treatment, animals showed a significant ($P < 0.05$, $n = 5$) increase in GABA as compared to control group. Ashwagandha (500 mg/kg) treatment showed a significant ($P < 0.01$, $n = 5$) increase in GABA as compared to alcohol consuming animals. While ashwagandha and shilajit in combination also showed significant ($P < 0.01$, $n = 5$) increase in GABA level in the brain as compared to the alcohol group. This significance was further increased ($P < 0.001$, $n = 5$) in diazepam treated group as compared to alcohol group [Figure 4].

**DISCUSSION**

In the present study, using conditioned place preference model, addiction to alcohol developed after 10 days of ethanol administration.
with animals spending 66% of time in ethanol-paired chamber over saline chamber. Chronic and excessive ethanol consumption followed by withdrawal results in abstinence syndrome. The predominant feature of alcohol withdrawal is anxiety, which is also considered to be the most important negative experience to revert to alcohol use. This may perturb central neurotransmitters and ion channel activity. Evidences indicate that during ethanol withdrawal, there is an upregulation of excitatory N-methyl-D-aspartate receptor (NMDA) receptors and a downregulation of inhibitory GABA<sub>A</sub> receptors. An important characteristic for most anti-addictive compounds is the elimination of withdrawal syndrome. Usually, a drug that either facilitates GABA action or decreases glutamate activity is effective against ethanol withdrawal-induced anxiety behavior. In our study, 5 days of abstinence postaddiction lead to the development of withdrawal anxiety. The anxiety reflected in reduction in time spent in open arm and number of open arm entries in EPM. Acute treatment with ashwagandha and shilajit or their combination reversed the ethanol withdrawal anxiety. The downregulation of GABA<sub>A</sub> receptor and/or decrease in the GABAergic transmission has been implicated in the withdrawal symptoms of ethanol. GABA mimetic effect of ashwagandha may prevent the declining GABA activity during alcohol withdrawal. Shilajit known for its anxiolytic and nootropic actions may increase neuronal dopamine levels partly responsible for its anxiolytic actions. Their amelioration of withdrawal anxiety was comparable to diazepam. Diazepam treatment also showed a significant increase in corticocerebellar gamma-aminobutyric acid levels. Values represent mean ± standard error of the mean n = 5

**Figure 4:** Effect of ashwagandha and shilajit on central nervous system neurotransmitter levels. (a) Changes in serotonin levels before and after treatment. Ashwagandha (500 mg/kg) as well as combined ashwagandha and shilajit treatment (day 21–30) lead to significant increase (P < 0.01) in corticocerebellar serotonin compared to untreated animals on ethanol (30 days). However, treatment with shilajit (50 mg/kg) alone or diazepam failed to increase serotonin levels compared to alcohol treated group. (b) Changes in dopamine levels before and after treatment. Shilajit (50 mg/kg) as well as ashwagandha and shilajit combination treatment (day 21–30) led to significant increase (P < 0.01) in corticocerebellar dopamine compared to untreated animals on ethanol (30 days). However, treatment with ashwagandha (500 mg/kg) alone or diazepam failed to increase serotonin levels compared to alcohol treated group. (c) Changes in gamma-aminobutyric acid levels before and after treatment. Ashwagandha (500 mg/kg) as well as combined ashwagandha and shilajit treatment (day 21–30) led to significant increase (P < 0.01) increase in corticocerebellar gamma-aminobutyric acid levels compared to untreated animals on ethanol (30 days). However, treatment with shilajit (50 mg/kg) alone failed to increase serotonin levels compared to alcohol-treated group. Diazepam treatment also showed a significant increase (P < 0.001) increase in corticocerebellar gamma-aminobutyric acid levels.
increased corticohippocampal levels of GABA. Decrease in GABAergic function after chronic administration of alcohol in experimental animals have been largely attributed to decrease in GABA<sub>G</sub> receptor expression and function. Ashwagandha treatment significantly increased GABA levels in the corticohippocampal lysates over both control and ethanol-treated animals. Ashwagandha has been previously reported to have GABA-mimetic activity by acting on both GABA<sub>A</sub> and GABA<sub>B</sub> receptors and may also have glycine mimetic action. Alcohol is an indirect GABA agonist. A plasma and CSF level of GABA is higher after initial withdrawal than after longer periods of abstinence. Diazepam, an established GABA agonist and anti-addictive compound, also showed increase in corticohippocampal GABA levels. Similar to previous reports, we also found an increase in serotonin levels in the corticohippocampal lysates of ethanol-exposed animals suggesting either excess release or reduced uptake of serotonin. Serotonergic system plays an important role in the ethanol intake, preference, and dependence via central mechanisms. Ashwagandha treatment showed an increase in cortical serotonin level over alcohol group. Others have shown that 5-HT<sub>1B</sub> receptor expression may also increase in animals receiving alcohol for several weeks. Increased serotonin activity at the 5-HT<sub>1B</sub> receptor by chronic alcohol exposure may also contribute to the alcohol withdrawal syndrome. Diazepam was also found to be the serotonin level in alcohol consuming brain. Alcohol consumption may induce a dopamine surge in the brain, turning it into a pleasurable experience thus reinforcing the action, and increasing consumption. Even low alcohol doses can increase dopamine release in parts of the nucleus accumbens. In contrast to other stimuli, alcohol-related stimuli maintain their motivational significance even after repeated alcohol administration, which may contribute to the craving for alcohol observed in alcoholics. In our study, chronic alcohol exposure increased corticohippocampal dopamine levels as compared to saline group while treatment with shilajit potentiated this effect further. Dopaminergic signal transmission may also be serotonin-mediated. 5-HT<sub>2</sub>A and 5-HT<sub>3</sub>A receptor signaling may stimulate dopaminergic neurons to release dopamine.

CONCLUSION

Here, we show ashwagandha and shilajit mediated dose-dependent decrease in ethanol withdrawal anxiety as well as a reduction in ethanol intake in mice. The primary mechanism responsible for anti-addictive activity of ashwagandha was found via GABAergic and serotonergic system while shilajit primarily mediated its effect by modulating dopamine levels in mice.

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Conflicts of interest

There are no conflicts of interest.

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